

# Package: MitoHEAR (via r-universe)

September 6, 2024

**Type** Package

**Title** Quantification of Mitochondrial DNA Heteroplasmy

**Version** 0.1.0

**Author** Gabriele Lubatti

**Maintainer** Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

**Description** R package that allows the estimation and downstream statistical analysis of the mitochondrial DNA Heteroplasmy calculated from single-cell datasets.

**License** GPL-3

**Depends** R (>= 4.0)

**Imports** Biostrings, circlize, ComplexHeatmap, dynamicTreeCut, GenomicRanges, ggplot2, gridExtra, IRanges, magrittr, mcclust, rdist, reshape2, rlist, Rsamtools,

**Suggests** clustree, fmsb, gam, karyoploteR, knitr, plotly, regioneR, rmarkdown, testthat

**VignetteBuilder** knitr

**biocViews** software

**Encoding** UTF-8

**LazyData** true

**Config/testthat/edition** 3

**RoxygenNote** 7.1.1

**BugReports** <https://github.com/ScialdoneLab/MitoHEAR/issues>

**Repository** <https://gabrielelubatti.r-universe.dev>

**RemoteUrl** <https://github.com/gabrielelubatti/mitohear>

**RemoteRef** HEAD

**RemoteSha** 3d655c46e42b82fc28fc0df066736adec37bb06a

## Contents

choose_features_clustering . . . . .	2
clustering_angular_distance . . . . .	4
detect_insertion . . . . .	5
dpt_test . . . . .	6
filter_bases . . . . .	7
get_distribution . . . . .	8
get_heteroplasmy . . . . .	8
get_raw_counts_allele . . . . .	10
get_wilcox_test . . . . .	11
plot_allele_frequency . . . . .	12
plot_base_coverage . . . . .	13
plot_batch . . . . .	14
plot_cells_coverage . . . . .	14
plot_condition . . . . .	15
plot_coordinate_cluster . . . . .	16
plot_coordinate_heteroplasmy . . . . .	16
plot_correlation_bases . . . . .	17
plot_distance_matrix . . . . .	18
plot_distribution . . . . .	19
plot_dpt . . . . .	19
plot_genome_coverage . . . . .	20
plot_heatmap . . . . .	21
plot_heteroplasmy . . . . .	22
plot_heteroplasmy_variability . . . . .	22
plot_spider_chart . . . . .	23
vi_comparison . . . . .	24
<b>Index</b>	<b>25</b>

---

choose\_features\_clustering  
*choose\_features\_clustering*

---

### Description

choose\_features\_clustering

### Usage

```
choose_features_clustering(  
  heteroplasmy_matrix,  
  allele_matrix,  
  cluster,  
  top_pos,  
  deepSplit_param,  
  minClusterSize_param,
```

```

    min_value_vector,
    threshold = 0.2,
    index,
    max_frac = 0.7
)

```

### Arguments

heteroplasmy_matrix	Third element returned by <i>get_heteroplasmy</i> .
allele_matrix	Fourth element returned by <i>get_heteroplasmy</i> .
cluster	Vector specifying a partition of the samples.
top_pos	Numeric value. Number of bases sorted with decreasing values of distance variance (see section <i>Details</i> below) among samples. If <i>relevant_bases</i> =NULL, then the bases for performing hierarchical clustering are the ones whose relative variance (variance of the base divided sum of variance among <i>top_pos</i> bases) is above <i>min_value</i> .
deepSplit_param	Integer value between 0 and 4 for the <i>deepSplit</i> parameter of the function <i>cutreeHybrid</i> . See section <i>Details</i> below.
minClusterSize_param	Integer value specifying the <i>minClusterSize</i> parameter of the function <i>cutreeHybrid</i> . See section <i>Details</i> below.
min_value_vector	Numeric vector. For each value in the vector, the function <i>clustering_angular_distance</i> is run with parameter <i>min_value</i> equal to one element of the vector <i>min_value_vector</i> .
threshold	Numeric value. If a base has heteroplasmy greater or equal to <i>threshold</i> in more than <i>max_frac</i> of cells, then the base is not considered for down stream analysis.
index	Fifth element returned by <i>get_heteroplasmy</i> .
max_frac	Numeric value. If a base has heteroplasmy greater or equal to <i>threshold</i> in more than <i>max_frac</i> of cells, then the base is not considered for down stream analysis.

### Value

Clustree plot.

### Author(s)

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

### See Also

<https://cran.r-project.org/package=clustree>

---

```
clustering_angular_distance
    clustering_angular_distance
```

---

## Description

For each pair of samples and for each base, an angular distance matrix is computed based on the four allele frequencies. Then only the angular distances corresponding to the `relevant_bases` are kept. If `relevant_bases` is NULL, then only the angular distances corresponding to the bases with relative distance variance among samples above `min_value` are kept. Finally the distance between each pair of samples is defined as the euclidean distance of the angular distances corresponding to the bases that pass the previous filtering step. On this final distance matrix, a hierarchical clustering approach is performed using the function `cutreeHybrid` of the package `dynamicTreeCut`.

## Usage

```
clustering_angular_distance(
  heteroplasmy_matrix,
  allele_matrix,
  cluster,
  top_pos,
  deepSplit_param,
  minClusterSize_param,
  threshold = 0.2,
  min_value,
  index,
  relevant_bases = NULL,
  max_frac = 0.7
)
```

## Arguments

<code>heteroplasmy_matrix</code>	Third element returned by <code>get_heteroplasmy</code> .
<code>allele_matrix</code>	Fourth element returned by <code>get_heteroplasmy</code> .
<code>cluster</code>	Vector specifying a partition of the samples.
<code>top_pos</code>	Numeric value. Number of bases sorted with decreasing values of distance variance (see section <i>Details</i> below) among samples. If <code>relevant_bases=NULL</code> , then the bases for performing hierarchical clustering are the ones whose relative variance (variance of the base divided sum of variance among <code>top_pos</code> bases) is above <code>min_value</code> .
<code>deepSplit_param</code>	Integer value between 0 and 4 for the <code>deepSplit</code> parameter of the function <code>cutreeHybrid</code> . See section <i>Details</i> below.

minClusterSize_param	Integer value specifying the <i>minClusterSize</i> parameter of the function <i>cutreeHybrid</i> . See section <i>Details</i> below.
threshold	Numeric value. If a base has heteroplasmy greater or equal to <i>threshold</i> in more than <i>max_frac</i> of cells, then the base is not considered for down stream analysis.
min_value	Numeric value. If <i>relevant_bases=NULL</i> , then the bases for performing hierarchical clustering are the ones whose relative variance (variance of the base divided sum of variance among <i>top_pos</i> bases) is above <i>min_value</i> .
index	Fifth element returned by <i>get_heteroplasmy</i> .
relevant_bases	Character vector of bases to consider as features for performing hierarchical clustering on samples. Default=NULL.
max_frac	Numeric value. If a base has heteroplasmy greater or equal to <i>threshold</i> in more than <i>max_frac</i> of cells, then the base is not considered for down stream analysis.

**Value**

It returns a list with 4 elements:

classification	Dataframe with two columns and <i>n_row</i> equal to <i>n_row</i> in <i>heteroplasmy_matrix</i> . The first column is the old cluster annotation provided by <i>cluster</i> . The second columns is the new cluster annotation obtained with hierarchical clustering on distance matrix based on heteroplasmy values.
dist_ang_matrix	Distance matrix based on heteroplasmy values as defined in the section <i>Details</i>
top_bases_dist	Vector of bases used for hierarchical clustering. If <i>relevant_bases</i> is not NULL, then <i>top_bases_dist=NULL</i>
common_idx	Vector of indices of samples for which hierarchical clustering is performed. If <i>index</i> is NULL, then <i>common_idx=NULL</i>

**Author(s)**

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

**See Also**

<https://www.rdocumentation.org/packages/dynamicTreeCut/versions/1.63-1/topics/cutreeHybrid>

---

detect\_insertion      *detect\_insertion*

---

**Description**

detect\_insertion

**Usage**

```
detect_insertion(ref_sequence, different_sequence, length_comparison = 10)
```

**Arguments**

- ref\_sequence     Character vector whose elements are the bases of a DNA sequence to use as reference.
- different\_sequence     Character vector whose elements are the bases of a DNA sequence different from the reference.
- length\_comparison     Integer number. Number of bases to consider for the comparison between the two DNA sequences in order to detect and remove insertions in the non-reference sequence.

**Value**

Character vector of the different\_sequence with length equal to ref\_sequence, after having removed the insertions.

**Author(s)**

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

---

dpt\_test

*dpt\_test*

---

**Description**

dpt\_test

**Usage**

```
dpt_test(heteroplasmy_matrix, time, index = NULL, method = "GAM")
```

**Arguments**

- heteroplasmy\_matrix     Third element returned by *get\_heteroplasmy*.
- time     Vector of diffusion pseudo time.
- index     index returned by *get\_heteroplasmy*.
- method     Character name denoting the method to choose for assigning an adjusted p value to each of the bases. Can be one of GAM, pearson and spearman. GAM: For each base, a GAM fit with formula  $z \sim \log(t)$  is performed between the heteroplasmy values (z) and the time (t). The p value from the table "Anova for Parametric Effects" is then assigned to the base. pearson,spearman:for each base, a pearson or spearman correlation test is performed between the heteroplasmy values and the time . The p value obtained from the test is then assigned to the base. In all the three possible methods, all the p values are then corrected with the method FDR.

**Value**

A data frame with 2 columns and number of rows equal to `n_col` in *heteroplasmy\_matrix*. In the first column there are the names of the bases while in the second column there are the adjusted p value.

**Author(s)**

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

**See Also**

<https://www.rdocumentation.org/packages/gam/versions/1.20/topics/gam>

---

filter_bases	<i>filter_bases</i>
--------------	---------------------

---

**Description**

filter\_bases

**Usage**

```
filter_bases(heteroplasmy_matrix, min_heteroplasmy, min_cells, index = NULL)
```

**Arguments**

heteroplasmy_matrix	Third element returned by <i>get_heteroplasmy</i> .
min_heteroplasmy	Numeric value.
min_cells	Numeric value.
index	Fifth element returned by <i>get_heteroplasmy</i> .

**Value**

Character vector of bases that have an heteroplasmy greater than *min\_heteroplasmy* in more than *min\_cells*.

**Author(s)**

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

---

```
get_distribution      get_distribution
```

---

### Description

```
get_distribution
```

### Usage

```
get_distribution(heteroplasmy_matrix, FUNCTION, index = NULL)
```

### Arguments

```
heteroplasmy_matrix      Third element returned by get_heteroplasmy.
FUNCTION                  A character specifying the function to be applied on each column of matrix. The
                          possible values are: mean,max,min,median and sum.
index                    index returned by get_heteroplasmy.
```

### Value

It returns a vector with length equal to `n_col` of *matrix* where each element contains the result of the operation defined by *FUNCTION*.

### Author(s)

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

---

```
get_heteroplasmy      get_heteroplasmy
```

---

### Description

It is one of the two main functions of the **MitoHEAR** package (together with *get\_raw\_counts\_allele*). It computes the allele frequencies and the heteroplasmy matrix starting from the counts matrix obtained with *get\_raw\_counts\_allele*.

### Usage

```
get_heteroplasmy(
  raw_counts_allele,
  name_position_allele,
  name_position,
  number_reads,
  number_positions,
  filtering = 1,
  my.clusters = NULL
)
```



**Arguments**

raw_counts_allele	A raw counts matrix obtained from <i>get_raw_counts_allele</i> .
name_position_allele	A character vector with elements specifying the genomic coordinate of the base and the allele (obtained from <i>get_raw_counts_allele</i> ).
name_position	A character vector with elements specifying the genomic coordinate of the base (obtained from <i>get_raw_counts_allele</i> ).
number_reads	Integer specifying the minimum number of counts above which we consider the base covered by the sample.
number_positions	Integer specifying the minimum number of bases that must be covered by the sample (with counts > <i>number_reads</i> ), in order to keep the sample for downstream analysis.
filtering	Numeric value equal to 1 or 2. If 1 then only the bases that are covered by all the samples are kept for the downstream analysis. If 2 then all the bases that are covered by more than 50% of the the samples in each cluster (specified by <i>my.clusters</i> ) are kept for the down-stream analysis. Default is 1.
my.clusters	Character vector specifying a partition of the samples. It is only used when filtering is equal to 2. Default is NULL

**Details**

Starting from *raw counts allele matrix*, the function performed two consequentially filtering steps. The first one is on the samples, keeping only the ones that cover a number of bases above *number\_positions*. The second one is on the bases, defined by the parameter *filtering*. The heteroplasmy for each sample-base pair is computed as  $1 - \max(f)$ , where  $f$  are the frequencies of the four alleles.

**Value**

It returns a list with 5 elements:

sum_matrix	A matrix (n_row=number of sample, n_col=number of bases) with the counts for each sample/base, for all the initial samples and bases included in the <i>raw counts allele matrix</i> .
sum_matrix_qc	A matrix (n_row=number of sample, n_col=number of bases) with the counts for each sample/base, for all the samples and bases that pass the two consequentially filtering steps.
heteroplasmy_matrix	A matrix with the same dimension of <i>sum_matrix_qc</i> where each entry (i,j) is the heteroplasmy for sample i in base j.
allele_matrix	A matrix (n_row=number of sample, n_col=4*number of bases) with allele frequencies, for all the samples and bases that pass the two consequentially filtering steps.
index	Indices of the samples that cover a base, for all bases and samples that pass the two consequentially filtering steps (if <i>filtering</i> = 2); if all the samples cover all the bases (that is the case for <i>filtering</i> = 1), then <i>index</i> is NULL

**Author(s)**

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

---

get\_raw\_counts\_allele *get\_raw\_counts\_allele*

---

**Description**

It is one the two main function of the **MitoHEAR** package (together with *get\_heteroplasmy*). The function allows to obtain a matrix of counts (n\_row = number of sample, n\_col= 4\*number of bases) of the four alleles in each base, for every sample. It takes as input a vector of sorted bam files (one bam file for each sample) and a fasta file for the genomic region of interest. It is based on the *pileup* function of the package Rsamtools.

**Usage**

```
get_raw_counts_allele(bam_input, path_fasta, cell_names, cores_number = 1)
```

**Arguments**

bam_input	Character vector of sorted bam files (full path). Each sample is defined by one bam file. For each bam file it is needed also the index bam file (.bai) at the same path.
path_fasta	Character string with full path to the fasta file of the genomic region of interest.
cell_names	Character vector of sample names.
cores_number	Number of cores to use.

**Value**

A list with three elements:

matrix\_allele\_counts

Matrix of counts (n\_row = number of sample, n\_col= 4\*number of bases) of the four alleles in each base, for every sample. The row names is equal to cell\_names.

name\_position\_allele

Character vector with length equal to n\_col of matrix\_allele\_counts. Each element specifies the coordinate of genomic position for a base and the allele.

name\_position

Character vector with length equal to n\_col of matrix\_allele\_counts. Each element specifies the coordinate of genomic position for a base.

**Author(s)**

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

**See Also**

<https://www.rdocumentation.org/packages/Rsamtools/versions/1.24.0/topics/pileup>

---

get_wilcox_test	<i>get_wilcox_test</i>
-----------------	------------------------

---

## Description

get\_wilcox\_test

## Usage

```
get_wilcox_test(heteroplasmy_matrix, cluster, label_1, label_2, index = NULL)
```

## Arguments

heteroplasmy_matrix	Third element returned by <i>get_heteroplasmy</i> .
cluster	Vector specifying a partition of the samples.
label_1	Character name of a first label included in cluster. It denotes the first group used for the Wilcoxon test
label_2	Character name of a second label included in cluster and different from label_1. it denotes the second group used for the Wilcoxon test.
index	Fifth element returned by <i>get_heteroplasmy</i> .

## Value

It returns a vector of length equal to n\_row in matrix. Each element stands for a base and it contains the adjusted p-value (FDR), obtained in unpaired two-samples Wilcoxon test from the comparison of the heteroplasmy between the label\_1 and label\_2 group.

## Author(s)

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

## See Also

<https://www.rdocumentation.org/packages/stats/versions/3.6.2/topics/wilcox.test>

---

plot\_allele\_frequency *plot\_allele\_frequency*

---

### Description

plot\_allele\_frequency

### Usage

```
plot_allele_frequency(  
  position,  
  heteroplasmy_matrix,  
  allele_matrix,  
  cluster,  
  names_allele_qc,  
  names_position_qc,  
  size_text,  
  index  
)
```

### Arguments

position	Character name of the base to plot.
heteroplasmy_matrix	Third element returned by <i>get_heteroplasmy</i> .
allele_matrix	Fourth element returned by <i>get_heteroplasmy</i> .
cluster	Vector specifying a partition of the samples.
names_allele_qc	Character vector with length equal to n_col of <i>allele_matrix</i> . Each element specifies the name of the base and the allele.
names_position_qc	Character vector with length equal to n_col of <i>allele_matrix</i> . Each element specifies the name of the base.
size_text	Character specifying the size of the text for <i>gridExtra</i> function <i>grid.arrange</i> )
index	Fifth element returned by <i>get_heteroplasmy</i> .

### Value

*grid.arrange* plot of allele frequencies of a specific base across samples divided according to cluster.

### Author(s)

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

### See Also

<https://cran.r-project.org/package=gridExtra>

---

plot\_base\_coverage     *plot\_base\_coverage*

---

## Description

plot\_base\_coverage

## Usage

```
plot_base_coverage(  
  sum_matrix,  
  sum_matrix_qc,  
  selected_cells,  
  interactive = FALSE,  
  text_size = 10  
)
```

## Arguments

`sum_matrix`     First element returned by the function *get\_heteroplasmy*.  
`sum_matrix_qc`   Second element returned by the function *get\_heteroplasmy*.  
`selected_cells`   Character vector with cells used fro plotting the coverage.  
`interactive`     Logical. If TRUE an interactive plot is produced.  
`text_size`       Character specifying the size of the text for ggplot2.

## Value

*ggplot2* object (if *interactive*=FALSE) or *plotly* object (if (if *interactive*=TRUE)

## Author(s)

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

## See Also

<https://plotly.com/r/>

---

plot_batch	<i>plot_batch</i>
------------	-------------------

---

**Description**

plot\_batch

**Usage**

```
plot_batch(position, heteroplasmy_matrix, batch, cluster, text_size, index)
```

**Arguments**

position	Character name of the base to plot.
heteroplasmy_matrix	Third element returned by <i>get_heteroplasmy</i> .
batch	Vector of batch names, with length equal to n_row of <i>heteroplasmy_matrix</i> .
cluster	Vector specifying a partition of the samples.
text_size	Character specifying the size of the text for ggplot2.
index	Fifth element returned by <i>get_heteroplasmy</i> .

**Value**

*ggplot2* object of the heteroplasmy level of a specific base across samples divided according to batch.

**Author(s)**

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

---

plot_cells_coverage	<i>plot_cells_coverage</i>
---------------------	----------------------------

---

**Description**

plot\_cells\_coverage

**Usage**

```
plot_cells_coverage(sum_matrix, cells_selected, cluster, interactive = FALSE)
```

**Arguments**

sum_matrix	First element returned by the function <i>get_heteroplasmy</i> .
cells_selected	Character vector of cells for which the coverage is computed.
cluster	Character vector with partition information for cells specified in <i>cells_selected</i>
interactive	Logical. If TRUE an interactive plot is produced.

**Value**

ggplot2 object (if *interactive*=FALSE) or plotly object (if *interactive*=TRUE)

**Author(s)**

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

**See Also**

<https://plotly.com/r/>

---

plot_condition	<i>plot_condition</i>
----------------	-----------------------

---

**Description**

plot\_condition

**Usage**

```
plot_condition(
  distribution_1,
  distribution_2,
  label_1,
  label_2,
  name_x,
  name_y,
  name_title
)
```

**Arguments**

distribution_1, distribution_2	Numeric vector
label_1	Character vector of length equal to distribution_1
label_2	Character vector of length equal to distribution_2
name_x	Character name specifying the xlab argument in <i>ggplot2</i> .
name_y	Character name specifying the ylab argument in <i>ggplot2</i> .
name_title	Character name specifying the ggtitle argument in <i>ggplot2</i> .

**Value**

*ggplot2* boxplot of the quantities specified by *distribution\_1* and *distribution\_2*, separated by the conditions denoted by *label\_1* and *label\_2*.

**Author(s)**

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

---

plot\_coordinate\_cluster  
*plot\_coordinate\_cluster*

---

**Description**

plot\_coordinate\_cluster

**Usage**

```
plot_coordinate_cluster(coordinate_dm, cluster)
```

**Arguments**

`coordinate_dm` Dataframe with samples on the rows and coordinates names on the columns.  
`cluster` Vector specifying a partition of the samples.

**Value**

*ggplot2* object.

**Author(s)**

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

---

plot\_coordinate\_heteroplasmy  
*plot\_coordinate\_heteroplasmy*

---

**Description**

plot\_coordinate\_heteroplasmy



**Usage**

```
plot_coordinate_heteroplasmy(
  coordinate_dm,
  heteroplasmy_matrix,
  index,
  name_base
)
```

**Arguments**

`coordinate_dm` Dataframe whit samples on the rows and coordinates names on the columns.  
`heteroplasmy_matrix` Third element returned by *get\_heteroplasmy*.  
`index` Fifth element returned by *get\_heteroplasmy*.  
`name_base` Character name specifying the base.

**Value**

*ggplot2* object.

**Author(s)**

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

---

plot\_correlation\_bases  
*plot\_correlation\_bases*

---

**Description**

plot\_correlation\_bases

**Usage**

```
plot_correlation_bases(bases_vector, index, heteroplasmy_matrix)
```

**Arguments**

`bases_vector` Character vector specifying the bases for which the spearman correlation across samples is computed.  
`index` Fifth element returned by *get\_heteroplasmy*.  
`heteroplasmy_matrix` Third element returned by *get\_heteroplasmy*.

**Value**

Heatmap plot produced by function *Heatmap*

**Author(s)**

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

**See Also**

<https://www.rdocumentation.org/packages/ComplexHeatmap/versions/1.10.2/topics/Heatmap>

---

plot\_distance\_matrix    *plot\_distance\_matrix*

---

**Description**

plot\_distance\_matrix

**Usage**

```
plot_distance_matrix(dist_ang_matrix, cluster)
```

**Arguments**

dist\_ang\_matrix

Distance matrix obtained from *clustering\_angular\_distance* (second element of the output).

cluster

Vector. Can be one of the two partitions returned by function *clustering\_angular\_distance* (first element of the output).

**Value**

Heatmap plot produced by function *Heatmap*

**Author(s)**

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

**See Also**

<https://www.rdocumentation.org/packages/ComplexHeatmap/versions/1.10.2/topics/Heatmap>

---

plot_distribution	<i>plot_distribution</i>
-------------------	--------------------------

---

**Description**

plot\_distribution

**Usage**

```
plot_distribution(quantity_counts_cell, name_x, name_title)
```

**Arguments**

quantity_counts_cell	Vector returned by <i>get_distribution</i>
name_x	Character name specifying the xlab argument in <i>ggplot2</i> .
name_title	Character name specifying the ggtitle argument in <i>ggplot2</i> .

**Value**

*ggplot2* density plot of the Vector quantity\_counts\_cell.

**Author(s)**

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

---

plot_dpt	<i>plot_dpt</i>
----------	-----------------

---

**Description**

plot\_dpt

**Usage**

```
plot_dpt(position, heteroplasmy_matrix, cluster, time, gam_fit_result, index)
```

**Arguments**

position	Character name of the base to plot.
heteroplasmy_matrix	Third element returned by <i>get_heteroplasmy</i> .
cluster	Vector specifying a partition of the samples.
time	Vector of diffusion pseudo time, with length equal to n_row of <i>heteroplasmy_matrix</i> .
gam_fit_result	Data frame returned by <i>dpt_test</i> .
index	Fifth element returned by <i>get_heteroplasmy</i> .

**Value**

ggplot object of the heteroplasmy level of a specific base across samples and the GAM fitted curve. The title shows the adjusted p value (FDR) for the position obtained from *get\_heteroplasmy*.

**Author(s)**

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

**See Also**

<https://cran.r-project.org/package=gam>

---

plot\_genome\_coverage    *plot\_genome\_coverage*

---

**Description**

plot\_genome\_coverage

**Usage**

```
plot_genome_coverage(biomart_file, path_fasta, chr_name, heteroplasmy_matrix)
```

**Arguments**

biomart_file	Character string with full path to the txt file downloaded from BioMart <a href="https://m.ensembl.org/info/data/biomart/index.html">https://m.ensembl.org/info/data/biomart/index.html</a> . It must have the following five columns: Gene.stable.ID, Gene.name, Gene.start..bp., Gene.end..bp., Chromosome.scaffold.name
path_fasta	Character string with full path to the fasta file of the genomic region of interest. It should be the same file used in <i>get_raw_counts_allele</i> .
chr_name	Character specifying the name of the chromosome of interest. It must be one of the names in the <i>Chromosome.scaffold.name</i> column from the <i>biomart_file</i> .
heteroplasmy_matrix	Third element returned by <i>get_heteroplasmy</i> .

**Value**

Plot as returned by *karyoploteR* package.

**Author(s)**

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

**See Also**

<http://bioconductor.org/packages/release/bioc/html/karyoploteR.html>

---

plot_heatmap	<i>plot_heatmap</i>
--------------	---------------------

---

**Description**

plot\_heatmap

**Usage**

```
plot_heatmap(
  new_classification,
  old_classification,
  dist_ang_matrix,
  cluster_columns = F,
  cluster_rows = T,
  name_legend
)
```

**Arguments**

new_classification	Character vector. Second column of the dataframe returned by function <i>clustering_angular_distance</i> (first element of the output).
old_classification	Character vector. First column of the dataframe returned by function <i>clustering_angular_distance</i> (first element of the output).
dist_ang_matrix	Distance matrix obtained from <i>clustering_angular_distance</i> (second element of the output).
cluster_columns	Logical. Parameter for cluster_columns argument of the function <i>Heatmap</i> in the package <i>ComplexHeatmap</i>
cluster_rows	Logical. Parameter for cluster_rows argument of the function <i>Heatmap</i>
name_legend	Character value. Parameter for name argument of the function <i>Heatmap</i>

**Value**

Heatmap plot produced by function *Heatmap*

**Author(s)**

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

**See Also**

<https://www.rdocumentation.org/packages/ComplexHeatmap/versions/1.10.2/topics/Heatmap>

---

plot\_heteroplasmy      *plot\_heteroplasmy*

---

**Description**

plot\_heteroplasmy

**Usage**

```
plot_heteroplasmy(position, heteroplasmy_matrix, cluster, index)
```

**Arguments**

position	Character name of the base to plot.
heteroplasmy_matrix	Third element returned by <i>get_heteroplasmy</i> .
cluster	Vector specifying a partition of the samples.
index	Fifth element returned by <i>get_heteroplasmy</i> .

**Value**

ggplot object of the heteroplasmy level of a specific base across samples divided according to cluster.

**Author(s)**

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

---

plot\_heteroplasmy\_variability  
    *plot\_heteroplasmy\_variability*

---

**Description**

plot\_heteroplasmy\_variability

**Usage**

```
plot_heteroplasmy_variability(  
  heteroplasmy_matrix,  
  cluster,  
  threshold = 0.1,  
  frac = FALSE,  
  index  
)
```

**Arguments**

heteroplasmy_matrix	Third element returned by <i>get_heteroplasmy</i> .
cluster	Vector specifying a partition of the samples.
threshold	Numeric value.
frac	Logical. If FALSE the absolute number of cells that have at least one base with heteroplasmy above <i>threshold</i> are shown separated by <i>cluster</i> . If TRUE, then the fraction of cells are shown.
index	Fifth element returned by <i>get_heteroplasmy</i> .

**Value**

*ggplot2* object

**Author(s)**

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

---

plot\_spider\_chart      *plot\_spider\_chart*

---

**Description**

plot\_spider\_chart

**Usage**

```
plot_spider_chart(name_base, cluster, heteroplasmy_matrix, index)
```

**Arguments**

name_base	Character name specifying the base.
cluster	Vector specifying a partition of the samples.
heteroplasmy_matrix	Third element returned by <i>get_heteroplasmy</i> .
index	Fifth element returned by <i>get_heteroplasmy</i> .

**Value**

radarchart plot produced by function *radarchart*.

**Author(s)**

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

**See Also**

<https://rdrr.io/cran/fmsb/man/radarchart.html>

---

vi_comparison	vi_comparison
---------------	---------------

---

### Description

We compute the variation of information (VI) between the partition provided by *new\_classification* and *old\_classification*. The VI between a random partitions (obtained with re-shuffle from original labels in *old\_classification*) and *old\_classification* is also computed. A distribution of VI values from random partitions is built. Finally, from the comparison with this distribution, an empirical p value is given to the VI of the unsupervised cluster analysis.

### Usage

```
vi_comparison(old_classification, new_classification, number_iter)
```

### Arguments

old_classification	Character vector. First column of the dataframe returned by function <i>clustering_angular_distance</i> (first element of the output).
new_classification	Character vector. Second column of the dataframe returned by function <i>clustering_angular_distance</i> (first element of the output).
number_iter	Integer value. Specify how many random partition are generated (starting from re-shuffle of labels in <i>old_classification</i> ).

### Value

Empirical p value.

### Author(s)

Gabriele Lubatti <gabriele.lubatti@helmholtz-muenchen.de>

### See Also

<https://www.rdocumentation.org/packages/mcclust/versions/1.0/topics/vi.dist>



# Index

choose\_features\_clustering, 2  
clustering\_angular\_distance, 4

detect\_insertion, 5  
dpt\_test, 6

filter\_bases, 7

get\_distribution, 8  
get\_heteroplasmy, 8  
get\_raw\_counts\_allele, 10  
get\_wilcox\_test, 11

plot\_allele\_frequency, 12  
plot\_base\_coverage, 13  
plot\_batch, 14  
plot\_cells\_coverage, 14  
plot\_condition, 15  
plot\_coordinate\_cluster, 16  
plot\_coordinate\_heteroplasmy, 16  
plot\_correlation\_bases, 17  
plot\_distance\_matrix, 18  
plot\_distribution, 19  
plot\_dpt, 19  
plot\_genome\_coverage, 20  
plot\_heatmap, 21  
plot\_heteroplasmy, 22  
plot\_heteroplasmy\_variability, 22  
plot\_spider\_chart, 23

vi\_comparison, 24